Research, original papers

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THE STRUCTURE AND CHEMICAL COMPOSITION OF THE FEMUR AFTER DIFFERENT METAL ALLOYS IMPLANTATION

ABSTRACT. Background. Osteointegration is a key factor for successful implant ingrowth. It depends on quality of bone, lack of initial stability, excessive loading, loosening or fracture of screw, and fracture of implant itself. Other factors that can affect osteointegration are implant composition and features of implant surface. Objective. The aim was to study the structure and chemical composition of an injured bone by scanning electron microscopy at different times after the implantation of metal alloys. Methods. Experiment was conducted on 90 rabbits, that were divided into 6 groups – 1 control and 5 experimental. We made round defect of femur in rabbits of control group and fill it by different alloys in experimental ones. Results. It was estimated that the implantation of TiVT6 and KTC-125 alloys in distal epiphysis leads to the formation of connective tissue around prosthesis and activation of bone remodeling. B- (Ti-Zr) alloy leads to early regression of the connective tissue around the implant and development of minimal changes in the structure and chemical composition of the bone during all periods of observation. Use of hydroxyapatite to coat metallic implants prevents connective capsule formation and stimulates bone growth around prosthesis that ensures its optimum fixation. Conclusion. Optimization of osteointegration leads to reduction of calcium loss. Hydroxyapatite coating of β- (Ti-Zr) alloy ensures the absence of stress-shielding displacement effect and development of bone micro-cracks.

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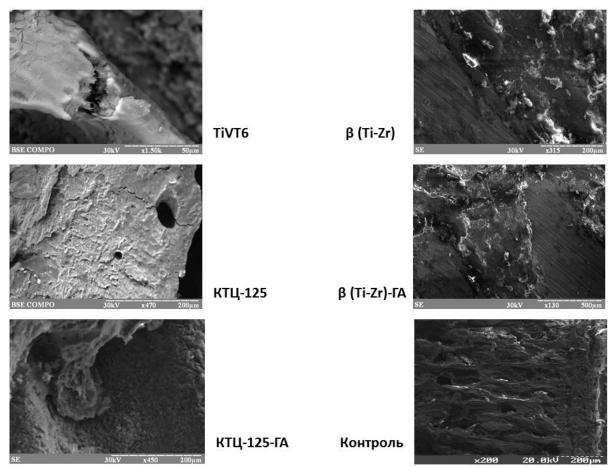


Fig. 1. Scanning electron microscopy of the area of bone around the implant 6 months after implantation of metals of different composition.

References

- 1. Frost HM. The Utah paradigm of skeletal physiology: an overview of its insights for bone, cartilage and collagenous tissue organs. J Bone Miner Metab. 2000;18(6):305-16. PMID: 11052462.
- 2. Albrektsson T, Wennerberg A. Oral implant surfaces: Part 1--review focusing on topographic and chemical properties of different surfaces and in vivo responses to them. Int J Prosthodont. 2004 SepOct;17(5):536-43. PMID: 15543910.
- 3. Anderson JM, Rodriguez A, Chang DT. Foreign body reaction to biomaterials. Semin Immunol. 2008 Apr;20(2):86-100. PMID: 18162407; PMCID: PMC2327202.
- 4. Joshi MG, Advani SG, Miller F, Santare MH. Analysis of a femoral hip prosthesis designed to reduce stress shielding. J Biomech. 2000 Dec;33(12):1655-62. PMID: 11006390.
- 5. Bai X, More K, Rouleau CM, Rabiei A.Functionally graded hydroxyapatite coatings doped with antibacterial components. Acta Biomater. 2010 Jun;6(6):2264-73. doi: 10.1016/j.actbio.2009.12.002. PMID: 19969112.

- 6. de Jonge LT, van den Beucken JJ, Leeuwenburgh SC, Hamers AA, Wolke JG, Jansen JA. In vitro responses to electrosprayed alkaline phosphatase/calcium phosphate composite coatings. Acta Biomater. 2009 Sep;5(7):2773-82. doi: 10.1016/j.actbio.2009.03.019. PMID: 19376000.
- 7. Dinda GP, Shin J, Mazumder J. Pulsed laser deposition of hydroxyapatite thin films on Ti-6Al-4V: effect of heat treatment on structure and properties. Acta Biomater. 2009 Jun;5(5):1821-30. doi: 10.1016/j.actbio.2009.01.027. PMID: 19269271.
- 8. Lamers E, Walboomers XF, Domanski M, te Riet J, van Delft FC, Luttge R, Winnubst LA, Gardeniers HJ, Jansen JA. The influence of nanoscale grooved substrates on osteoblast behavior and extracellular matrix deposition. Biomaterials. 2010 Apr;31(12):3307-16. doi: 10.1016/j.biomaterials.2010.01.034. PMID: 20122723.
- 9. Mendonça G, Mendonça DB, Aragão FJ, Cooper LF. Advancing dental implant surface technology--from micron- to nanotopography. Biomaterials. 2008 Oct;29(28):3822-35. doi: 10.1016/j.biomaterials.2008.05.012. PMID: 18617258.
- 10. Ramakrishna S, Mayer J, Wintermantel E, Leong KW. Biomedical applications of polymercomposite materials: A review. Compos Sci Technol. 2001;61:1189-224.
- 11. Schliephake H., Scharnweber D. Chemical and biological functionalization of titanium for dental implants. J Mater Chem. 2008;18:2404-14.
- 12. Zhao G, Schwartz Z, Wieland M, Rupp F, Geis-Gerstorfer J, Cochran DL, Boyan BD. High surface energy enhances cell response to titanium substrate microstructure. J Biomed Mater Res A. 2005 Jul 1;74(1):49-58. PMID: 15924300.